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cis-Golgi Marker

Anti-GM130 mAb

Code No.	Clone	Subclass	Quantity	Concentration
M179-3MS	5G8	Mouse IgG2a κ	20 μ L	1 mg/mL

BACKGROUND: The Golgi apparatus is a eukaryotic organelle, which is mainly devoted to processing the proteins synthesized in the endoplasmic reticulum (ER). GM130 is a member of the golgin family of coiled-coil proteins that localizes predominantly to the cis-Golgi. GM130 might participate in ER-Golgi traffic.

SOURCE: This antibody was purified from hybridoma (clone 5G8) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell P3U1 with C3H mouse lymphocyte immunized with the synthetic peptide corresponding to C-terminal of human GM130.

FORMULATION: 20 μ g IgG in 20 μ L volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at -20°C .

REACTIVITY: This antibody reacts with human GM130 on Western blotting, Immunoprecipitation, and Immunocytochemistry.

APPLICATIONS:

Western blotting: 1 μ g/mL for chemiluminescence detection system

Immunoprecipitation: 2 μ g/300 μ L of cell extract from 3×10^6 cells

Immunohistochemistry: Not tested

Immunocytochemistry: 5 μ g/mL

Flow cytometry: Not tested

Detailed procedure is provided in the following **PROTOCOLS**.

SPECIES CROSS REACTIVITY:

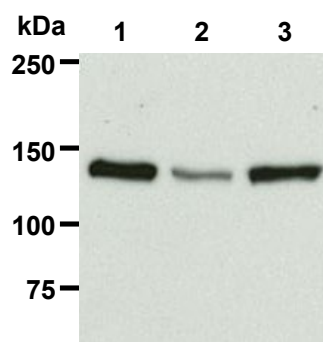
Species	Human	Mouse	Rat
Cells	Hela, 293T A549	Not Tested	Not Tested
Reactivity on WB	+		

INTENDED USE:

For Research Use Only. Not for use in diagnostic procedures.

REFERENCES:

- 1) Diao, A., *et al.*, *J. Biol. Chem.* **283**, 6957-6967 (2008)
- 2) Alvarez, C., *et al.*, *J. Biol. Chem.* **276**, 2693-2700 (2001)



Western blot analysis of GM130 in HeLa (1), 293T (2) and A549 (3) using M179-3.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

PROTOCOLS:

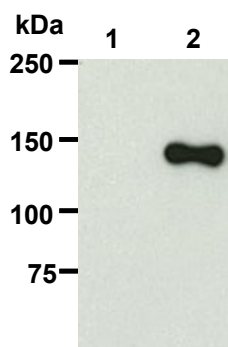
SDS-PAGE & Western Blotting

- 1) Wash cells (approximately 1×10^7 cells) 3 times with PBS and resuspend them in 1 mL of Laemmli's sample buffer.
- 2) Boil the samples for 3 minutes and centrifuge. Load 20 μ L of sample per lane on a 1-mm-thick SDS-polyacrylamide gel and carry out electrophoresis.
- 3) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm² for 1 hour in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacturer's manual for precise transfer procedure.
- 4) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature, or overnight at 4°C .
- 5) Incubate the membrane for 1 hour at room temperature with primary antibody diluted with PBS (pH 7.2) containing 1% skimmed milk as suggested in the **APPLICATIONS**. (The concentration of antibody will depend on the conditions.)
- 6) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 minutes x 3 times).
- 7) Incubate the membrane with 1:10,000 HRP-conjugated anti-mouse IgG (MBL; code no. 330) diluted with 1%

skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.

- 8) Wash the membrane with PBS-T (5 minutes x 3 times).
- 9) Wipe excess buffer off the membrane, and incubate membrane with an appropriate chemiluminescence reagent for 1 minute.
- 10) Remove extra reagent from the membrane by dabbing with a paper towel, and seal it in plastic wrap.
- 11) Expose the membrane onto an X-ray film in a dark room for 6 minutes. Develop the film under usual settings. The conditions for exposure and development may vary.

(Positive controls for Western blotting; HeLa, 293T, A549)



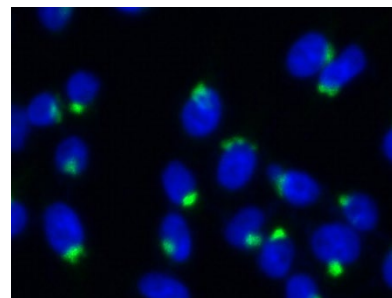
Immunoprecipitation of GM130 from HeLa with mouse IgG2a isotypic control, M076-3 (1) or M179-3 (2). After immunoprecipitated with the antibody, immunocomplexes were resolved on SDS-PAGE and immunoblotted with PM061.

Immunoprecipitation

- 1) Wash cells (approximately 1×10^7 cells) 3 times with PBS and resuspend them in 1 mL of cold Lysis buffer (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.05% NP-40) containing protease inhibitors at appropriate concentrations. Incubate it at 4°C with rotating for 30 minutes; thereafter, briefly sonicate the mixture (up to 20 seconds).
- 2) Centrifuge the tube at 12,000 x g for 10 minutes at 4°C and transfer the supernatant to another fresh tube.
- 3) Add primary antibody as suggested in the **APPLICATIONS** into 300 μ L of the supernatant. Mix well and incubate with gentle agitation for 30-120 minutes at 4°C. Add 20 μ L of 50% protein A agarose beads resuspended in the cold Lysis buffer. Mix well and incubate with gentle agitation for 60 minutes at 4°C.
- 4) Wash the beads 3-5 times with the cold Lysis buffer (centrifuge the tube at 2,500 x g for 10 seconds).
- 5) Resuspend the beads in 20 μ L of Laemmli's sample buffer, boil for 3-5 minutes, and centrifuge for 5 minutes. Use 10 μ L/lane for the SDS-PAGE analysis.

(See **SDS-PAGE & Western blotting.**)

(Positive control for Immunoprecipitation; HeLa)



Immunocytochemical detection of GM130 in HeLa using M179-3.

Green: anti-GM130

Blue: DAPI

Immunocytochemistry

- 1) Culture the cells in the appropriate condition on a glass slide. (for example, spread 1×10^4 cells of HeLa cells for one slide, then incubate in a CO₂ incubator for one night.)
- 2) Wash the cells 3 times with PBS.
- 3) Fix the cells by immersing the slide in PBS containing 4% paraformaldehyde for 10 minutes at room temperature.
- 4) Wash the glass slide 3 times with PBS.
- 5) Immerse the slide in PBS containing 0.2% Triton X-100 for 10 minutes at room temperature.
- 6) Wash the glass slide 2 times with PBS.
- 7) Add the primary antibody diluted with 2% FCS/PBS as suggested in the **APPLICATIONS** onto the cells and incubate for 30 minutes at room temperature (Optimization of antibody concentration or incubation condition are recommended if necessary.)
- 8) Wash the glass slide 2 times with PBS.
- 9) Add 100 μ L of 1:500 Alexa Fluor[®]488 conjugated anti-mouse IgG (Invitrogen; code no. 53818A) diluted with PBS onto the cells. Incubate for 30 minutes at room temperature. Keep out light by aluminum foil.
- 10) Wash the glass slide 2 times with PBS.
- 11) Counter stain with DAPI for 5 minutes at room temperature.
- 12) Wash the glass slide 2 times with PBS.
- 13) Wipe excess liquid off the slide but take care not to touch the cells. Never leave the cells to dry.
- 14) Promptly add mounting medium onto the slide, then put a cover slip on it.

(Positive control for Immunocytochemistry; HeLa)

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