For Research Use Only.
Not for use in diagnostic procedures.

MBL
A JSR Life Sciences Company

Smart-IP Series

Anti-HA-tag mAb-Magnetic Beads

CODE No.	M132-11
CLONALITY	Monoclonal
CLONE	5D8
ISOTYPE	Mouse IgG1 κ
QUANTITY	20 tests (Slurry: 1 mL)
SOURCE	Purified IgG from hybridoma supernatant
IMMUNOGEN	Carrier protein conjugated synthetic peptide, YPYDVPDYA (HA-tag)
REACTIVITY	This antibody reacts with N-terminal and C-terminal HA-tagged proteins.
FORMULATION	10 mg magnetic beads in 1 mL PBS/0.1% BSA/0.1% ProClin 150
STORAGE	This beads suspension is stable for one year from the date of purchase when stored at 4°C. If bead agglomeration is observed, please disperse the agglomerations by careful pipetting. *In particular, please check the inner wall of the vial and cap.

APPLICATION-CONFIRMED

Immunoprecipitation 50 µL of beads slurry/sample

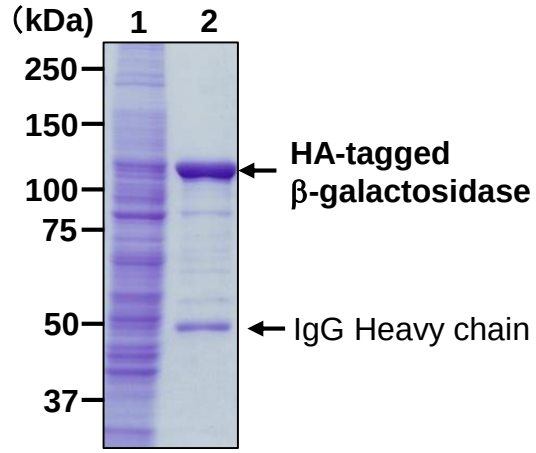
*The purification capacity of Anti-HA-tag mAb-Magnetic Beads varies depending upon the characteristics of a HA-tagged protein. For example, 50 µL of beads slurry bounds 3.5 µg of a HA-tagged protein (35 kDa).

For more information, please visit our website **https://ruo.mbl.co.jp**.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

Immunoprecipitation

1) Wash 1 x 10^7 cells 3 times with PBS and suspend with 1 mL of cold Lysis buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1% NP-40] containing appropriate protease inhibitors.
2) Centrifuge the tube at 12,000 x g for 5 min. at 4°C and transfer the supernatant to another tube.
3) Add magnetic beads as suggested in the **APPLICATION** into 300 µL of the supernatant prepared in step 2). Mix well and incubate with gentle agitation for 1 hr. at 4°C.
4) Place the tube on the magnetic rack (MBL; code no. 3190) for a few seconds.
5) Remove the supernatant.
6) Wash the beads 4 times with 1 mL of cold Wash buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 0.05% NP-40] (place the tube on the magnetic rack for a few seconds).
7) Resuspend the magnetic beads in 20 µL of Laemmli's sample buffer, boil for 5 min., and place the tube on the magnetic rack for a few seconds.
8) Load 20 µL of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel and carry out electrophoresis.
9) Visualize the protein bands by CBB staining.



Immunoprecipitation of HA-tagged protein

Sample: HA-tagged β-galactosidase/293T whole cell lysate
Lane 1: Input (10 µL/lane)
Lane 2: Post-IP beads of Anti-HA-tag mAb (MBL; code no. M132-11)