

MONOCLONAL ANTIBODY

Mouse IgM (isotype control)

Code No.	Clone	Subclass	Quantity	Concentration
M079-3	7E10	Mouse IgM	100 μ L	1 mg/mL

SOURCE: This antibody was purified from hybridoma (clone 7E10) supernatant by affinity chromatography. This hybridoma was established by fusion of mouse myeloma cell P3U1 with Balb/c mouse lymphnodes immunized with KLH.

FORMULATION: 100 μ g IgM in 100 μ L volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at -20°C .

REACTIVITY: No specific binding detected on human peripheral blood leukocytes.

APPLICATION:

Flow cytometry: This antibody can be used as a negative isotypic control. The concentration to be used will be dependent on condition.

Detailed procedure is provided in the following **PROTOCOLS**.

INTENDED USE:

For Research Use Only. Not for use in diagnostic procedures.

PROTOCOLS:

Flow cytometric analysis for floating cells

We usually use Fisher tubes or equivalents as reaction tubes for all step described below.

- 1) Wash the cells 3 times with washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.09% NaN_3].
*Azide may react with copper or lead in plumbing system to form explosive metal azides. Therefore, always flush plenty of water when disposing materials containing azide into drain.
- 2) Resuspend the cells with washing buffer (5×10^6 cells/mL).
- 3) Add 50 μ L of the cell suspension into each tube, and centrifuge at 500 x g for 1 minute at room temperature ($20\sim 25^{\circ}\text{C}$). Remove supernatant by careful aspiration.
- 4) Add 10 μ L of Clear Back (Human Fc receptor blocking reagent, MBL; code no. MTG-001) to the cell pellet after tapping. Mix well and incubate for 5 minutes at room temperature.

- 5) Add 40 μ L of the mouse IgM isotype control (25 $\mu\text{g/mL}$) diluted with the washing buffer. Mix well and incubate for 30 minutes at room temperature.
- 6) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 7) Add 20 μ L of 1:100 FITC conjugated anti-mouse IgM (Beckman coulter; code no. IM0821) diluted with the washing buffer. Mix well and incubate for 20 minutes at room temperature.
- 8) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 9) Resuspend the cells with 500 μ L of the washing buffer and analyze by a flow cytometer.

Flow cytometric analysis for whole blood cells

We usually use Falcon tubes or equivalents as reaction tubes for all step described below.

- 1) Add 50 μ L of the mouse IgM isotype control (20 $\mu\text{g/mL}$) diluted with washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.09% NaN_3] into each tube.
- 2) Add 50 μ L of whole blood into each tube. Mix well and incubate for 30 minutes at room temperature ($20\sim 25^{\circ}\text{C}$).
- 3) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 4) Add 20 μ L of secondary antibody 1:100 FITC conjugated anti-mouse IgM (Beckman coulter; code no. IM0821) diluted with the washing buffer. Mix well and incubate for 15 minutes at room temperature.
- 5) Lyse with OptiLyse C (for analysis on Beckman Coulter instruments) or OptiLyse B (for analysis on BD instruments), using the procedure recommended in the respective package inserts.
- 6) Add 1 mL of H_2O to each tube and incubate for 10 minutes at room temperature.
- 7) Centrifuge at 500 x g for 1 minute at room temperature.
- 8) Add 1 mL of washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 9) Resuspend the cells with 500 μ L of the washing buffer and analyze by a flow cytometer.

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