**For Research Use Only.**
**Not for use in diagnostic procedures.**

*Smart-IP Series*

# Anti-Myc-tag mAb-Magnetic Beads

**CODE No.** M047-11

**CLONALITY** Monoclonal
**CLONE** PL14
**ISOTYPE** Mouse IgG1
**QUANTITY** 20 tests (Slurry: 1 mL)

**SOURCE** Purified IgG from mouse ascites fluid
**IMMUNOGEN** 6myc-tagged fusion protein
**FORMULATION** 15 mg magnetic beads in 1 mL PBS/0.1% BSA/0.09% NaN3
*Azide may react with copper or lead in plumbing system to form explosive metal azides. Therefore, always flush plenty of water when disposing materials containing azide into drain.
**STORAGE** This beads suspension is stable for one year from the date of purchase when stored at 4°C.

**APPLICATION-CONFIRMED**
Immunoprecipitation 50 µL of beads slurry/sample
*The purification capacity of Anti-Myc-tag mAb-Magnetic Beads varies depending upon the characteristics of a Myc-tagged protein. For example, 50 µL of beads slurry bounds 0.4 µg of a Myc-tagged protein (35 kDa).

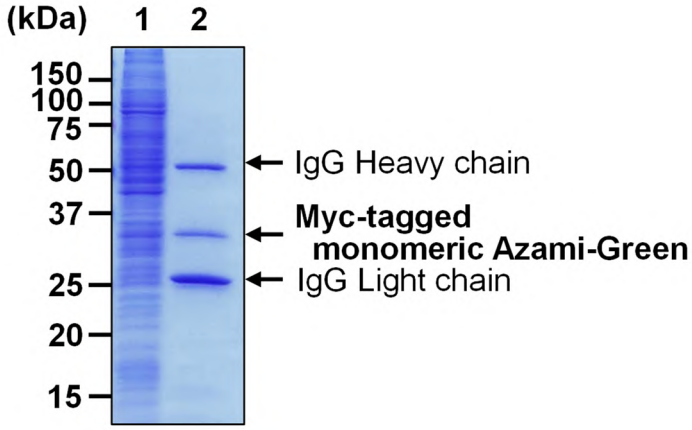
**REFERENCE**
1) Jain, P., *et al., Oncogene* **36**, 6348-6358 (2017) [Co-IP]

For more information, please visit our website at **https://ruo.mbl.co.jp/**.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

## Immunoprecipitation

1) Wash 1 x $10^7$ cells 3 times with PBS and suspend with 1 mL of cold Lysis buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1% NP-40] containing appropriate protease inhibitors.
2) Centrifuge the tube at 12,000 x g for 5 min. at 4°C and transfer the supernatant to another tube.
3) Add magnetic beads as suggested in the **APPLICATION** and purified Myc-tagged protein into 300 µL of the supernatant prepared in step 2). Mix well and incubate with gentle agitation for 1 hr. at 4°C.
4) Place the tube on the magnetic rack (MBL, code no. 3190) for a few seconds.
5) Remove the supernatant.
6) Wash the beads 4 times with 1 mL of cold Wash buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 0.05% NP-40] (place the tube on the magnetic rack for a few seconds).
7) Resuspend the magnetic beads in 20 µL of Laemmli's sample buffer, boil for 2 min., and place the tube on the magnetic rack for a few seconds.
8) Load 20 µL of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel and carry out electrophoresis.
9) Visualize the protein bands by CBB staining.




### *Immunoprecipitation of Myc-tagged protein*

Sample: 293T cell lysate from 3x$10^6$ cells + Myc-tagged monomeric Azami-Green 10 µg
Lane 1: Input (5 µL/lane)
Lane 2: Post-IP beads of Anti-Myc-tag mAb (MBL, code no. M047-11)