



For Research Use Only, Not for use in diagnostic procedures

ELISA Kit for Measuring Human REDD1

# CircuLex Human REDD1 ELISA Kit

Cat# CY-8113

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## Intended Use

The MBL Research Product **CircuLex Human REDD1 ELISA Kit** is used for the quantitative measurement of human REDD1 in cell lysate.

Individual users should determine appropriate conditions when using other types of samples.

**This assay kit is for research use only and not for use in diagnostic or therapeutic procedures.**

## Storage

- Upon receipt store all components at 4°C.
- Don't expose reagents to excessive light.

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## Introduction

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REDD1 (Regulated in Development and DNA Damage 1) (1) is also called DNA Damage-inducible Transcript 4 Protein, HIF-1 Responsive Protein RTP801 (2) and Dexamethasone-induced gene 2 (3). REDD1 is rapidly induced by a wide variety of stress conditions; arsenic, hypoxia, dexamethasone, thapsigargin, tunicamycin and heat shock and DNA damages; ionizing radiation, ultraviolet radiation, DNA alkylation. REDD1 is broadly expressed in most adult tissues, with lowest levels in brain, skeletal muscle and intestine and is upregulated in substantia nigra neurons from Parkinson disease patients (4) as well as in alveolar septa of lungs of individuals with advanced emphysema compared with normal lungs (5).

REDD1 regulates cell growth, proliferation and survival via inhibition of the activity of mTORC by stabilizing the TSC1-TSC2 inhibitory complex and enhances oxidative stress-dependent cell death (6, 7). REDD1 inhibits neuronal differentiation and neurite outgrowth mediated by NGF via its effect on mTORC1 activity.

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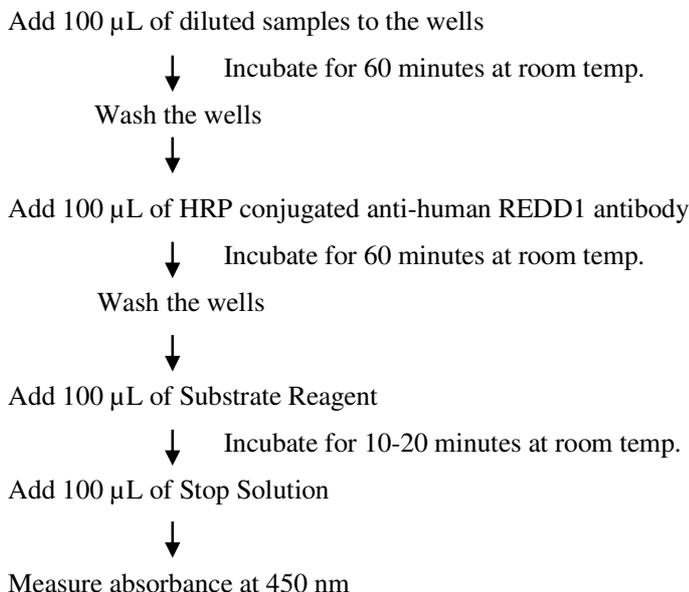
## Principle of the Assay

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The MBL Research Product **CircuLex Human REDD1 ELISA Kit** employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for human REDD1 is pre-coated onto a microplate. Standards and samples are pipetted into the wells and the immobilized antibody binds any human REDD1 present. After washing away any unbound substances, an HRP conjugated antibody specific for human REDD1 is added to the wells. Following a wash to remove any unbound antibody HRP conjugate, the remaining conjugate is allowed to react with the substrate H<sub>2</sub>O<sub>2</sub>-tetramethylbenzidine. The reaction is stopped by addition of acidic solution and absorbance of the resulting yellow product is measured at 450 nm. The absorbance is proportional to the concentration of human REDD1. A standard curve is constructed by plotting absorbance values versus human REDD1 concentrations of calibrators, and concentrations of unknown samples are determined using this standard curve.

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## Summary of Procedure



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## Materials Provided

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All samples and standards should be assayed in duplicate. The following components are supplied and are sufficient for the one 96-well microplate kit.

**Microplate:** One microplate supplied ready to use, with 96 wells (12 strips of 8-wells) in a foil, zip-lock bag with a desiccant pack. Wells are coated with anti-human REDD1 antibody as a capture antibody.

**10X Wash Buffer:** One bottle containing 100 mL of 10X buffer containing Tween®-20.

**Dilution Buffer:** One bottle containing 50 mL of 1X buffer; use for reconstitution of Human REDD1 Standard and sample dilution. Ready to use.

**Human REDD1 Standard:** One vial containing X\* ng of lyophilized recombinant human REDD1.

**\*The amount is changed depending on lot. See the real "User's Manual" included in the kit box.**

**HRP conjugated Detection Antibody:** One bottle containing 12 mL of HRP (horseradish peroxidase) conjugated anti-human REDD1 antibody. Ready to use.

**Substrate Reagent:** One bottle containing 20 mL of the chromogenic substrate, tetra-methylbenzidine (TMB). Ready to use.

**Stop Solution:** One bottle containing 20 mL of 1 N H<sub>2</sub>SO<sub>4</sub>. Ready to use.

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## Materials Required but not Provided

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- **Pipettors:** 2-20  $\mu$ L, 20-200  $\mu$ L and 200-1,000  $\mu$ L precision pipettors with disposable tips.
- **Precision repeating pipettor**
- **Orbital microplate shaker**
- **Microcentrifuge and tubes** for sample preparation.
- **Vortex mixer**
- **(Optional) Microplate washer:** Manual washing is possible but not preferable.
- **Plate reader** capable of measuring absorbance in 96-well plates at dual wavelengths of 450 nm/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. The plate can also be read at a single wavelength of 450 nm, which will give a somewhat higher reading.
- **(Optional) Software package facilitating data generation and analysis**
- **500 or 1,000 mL graduated cylinder.**
- **Reagent reservoirs**
- **Deionized water of the highest quality**
- **Disposable paper towels**

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## Precautions and Recommendations

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- **Although we suggest to conduct experiments as outlined below, the optimal experimental conditions will vary depending on the parameters being investigated, and must be determined by the individual user.**
- Allow all the components to come to room temperature before use.
- All microplate strips that are not immediately required should be returned to the zip-lock pouch, which must be carefully resealed to avoid moisture absorption.
- Do not use kit components beyond the indicated kit expiration date.
- Use only the microtiter wells provided with the kit.
- Rinse all detergent residue from glassware.
- Use deionized water of the highest quality.
- Do not mix reagents from different kits.
- The buffers and reagents in this kit may contain preservatives or other chemicals. Care should be taken to avoid direct contact with these reagents.
- Do not mouth pipette or ingest any of the reagents.
- Do not smoke, eat, or drink when performing the assay or in areas where samples or reagents are handled.
- Dispose of tetra-methylbenzidine (TMB) containing solutions in compliance with local regulations.
- Avoid contact with the acidic Stop Solution and Substrate Solution, which contains hydrogen peroxide.
- Wear gloves and eye protection when handling immunodiagnostic materials and samples of human origin, and these reagents. In case of contact with the Stop Solution and the Substrate Solution, wash skin thoroughly with water and seek medical attention, when necessary.
- **Biological samples may be contaminated with infectious agents. Do not ingest, expose to open wounds or breathe aerosols. Wear protective gloves and dispose of biological samples properly.**
- **CAUTION: Sulfuric Acid is a strong acid. Wear disposable gloves and eye protection when handling Stop Solution.**

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## Sample Collection and Storage

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**Cell lysate:** Several extraction methods can be used for measurement cellular proteins. The following protocol is provided as an example of suitable methods. All steps of cell lysate preparation should be performed at 4°C.

1. Harvest and pellet cells by centrifugation using standard methods.
2. Resuspend the cell pellet with the cell lysis buffer (25 mM Tris-HCl, pH 7.5, 250 mM NaCl, 0.1 % NP-40, 1 mM EDTA, 0.2 mM PMSF, 1 µg/mL pepstatin, 0.5 µg/mL leupeptin, 0.2 mM DTT).
3. Lyse the resuspended cells using either a Dounce homogenizer, sonication, or three cycles of freezing and thawing.
4. Transfer extracts to microcentrifuge tubes and centrifuge at 15,000 rpm for 10 minutes at 4°C.
5. Aliquot cleared lysate to a clean microfuge tube.
6. Assay immediately or store the samples on ice for a few hours before assaying. Aliquots of the samples may also be stored below -70°C for extended periods of time. Avoid repeated freeze-thaw cycles.

**Note-1:** The above procedures are intended only as a guideline. The optimal experimental conditions will vary depending on the parameters being investigated, and must be determined by the individual user.

**Note-2:** Individual users should determine appropriate conditions when using other types of samples which doesn't be indicated in this manual.

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## Detailed Protocol

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The MBL Research Product **CircuLex Human REDD1 ELISA Kit** is provided with removable strips of wells so the assay can be carried out on separate occasions using only the number of strips required for the particular determination. Since experimental conditions may vary, an aliquot of the Human REDD1 Standard within the kit should be included in each assay as a calibrator. Disposable pipette tips and reagent troughs should be used for all liquid transfers to avoid cross-contamination of reagents or samples.

### Preparation of Working Solutions

All reagents need to be brought to room temperature prior to the assay. Assay reagents are supplied ready-to-use, with the exception of **10X Wash Buffer** and **Human REDD1 Standard**.

1. Prepare a working solution of Wash Buffer by adding 100 mL of the **10X Wash Buffer** to 900 mL of deionized (distilled) water. Mix well. Store at 4°C for two weeks or -20°C for long-term storage.
2. Reconstitute **Human REDD1 Standard** with **X\*** mL of **Dilution Buffer** by gently mixing. After reconstitution, immediately dispense it in small aliquots (e.g. 100 µL) to plastic micro-centrifuge tubes and store below -70°C to avoid non-specific adsorption to glass surface and multiple freeze-thaw cycles. The concentration of the reconstituted Human REDD1 Standard should be **128 ng/mL**, which is referred to as the **Master Standard** of human REDD1.

**\*The volume is changed depending on lot. See the real "User's Manual" included in the kit box.**

Prepare Standard Solutions as follows:

Use the **Master Standard** to produce a dilution series (below). Mix each tube thoroughly before the next transfer. **Std.1 (12.8 ng/mL)** serves as the highest standard. The **Dilution Buffer** serves as the zero standard (Blank).

	Volume of Standard	Dilution Buffer	Concentration
Std.1	60 µL of Master Standard (128 ng/mL)	540 µL	12.8 ng/mL
Std.2	300 µL of Std. 1 (12.8 ng/mL)	300 µL	6.4 ng/mL
Std.3	300 µL of Std. 2 (6.4 ng/mL)	300 µL	3.2 ng/mL
Std.4	300 µL of Std. 3 (3.2 ng/mL)	300 µL	1.6 ng/mL
Std.5	300 µL of Std. 4 (1.6 ng/mL)	300 µL	0.8 ng/mL
Std.6	300 µL of Std. 5 (0.8 ng/mL)	300 µL	0.4 ng/mL
Std.7	300 µL of Std. 6 (0.4 ng/mL)	300 µL	0.2 ng/mL
Blank	-	300 µL	0 ng/mL

**Note:** Do not use a Repeating pipette. Change tips for every dilution. Wet tip with Dilution Buffer before dispensing.

### Sample Preparation

Dilute samples with **Dilution Buffer**.

- Samples, e.g. cell lysates and tissue extracts, require the proper dilution ratio. Dilution ratio varies depending on the sample types and the isolation and extraction conditions.

## Assay Procedure

1. Remove the appropriate number of microtiter wells from the foil pouch and place them into the well holder. Return any unused wells to the foil pouch, refold, seal with tape and store at 4°C.
2. Dilute samples with **Dilution Buffer**. (See "Sample Preparation" above.)
3. Pipette **100 µL** of **Standard Solutions (Std1-Std7, Blank)** and **diluted samples** in duplicates, into the appropriate wells.
4. Incubate the plate **at room temperature (ca.25°C) for 60 minutes**, shaking at ca. 300 rpm on an orbital microplate shaker.
5. Wash 4-times by filling each well with Wash Buffer (350 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer.
6. Add **100 µL** of **HRP conjugated Detection Antibody** into each well.
7. Incubate the plate **at room temperature (ca.25°C) for 60 minutes**, shaking at ca. 300 rpm on an orbital microplate shaker.
8. Wash 4-times by filling each well with Wash Buffer (350 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer.
9. Add **100 µL** of **Substrate Reagent**. Avoid exposing the microtiter plate to direct sunlight. Covering the plate with e.g. aluminum foil is recommended. Return Substrate Reagent to 4°C immediately after the necessary volume is removed
10. Incubate the plate **at room temperature (ca.25°C) for 10-20 minutes**, shaking at ca. 300 rpm on an orbital microplate shaker. The incubation time may be extended up to 30 minutes if the reaction temperature is below 20°C.
11. Add **100 µL** of **Stop Solution** to each well in the same order as the previously added Substrate Reagent.
12. Measure absorbance in each well using a spectrophotometric microplate reader at dual wavelengths of 450/540 nm. Dual wavelengths of 450/550 or 450/595 nm can also be used. Read the microplate at 450 nm if only a single wavelength can be used. Wells must be read within 30 minutes of adding the Stop Solution.

**Note-1:** Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

**Note-2:** Reliable standard curves are obtained when either O.D. values do not exceed 0.25 units for the blank (zero concentration), or 3.0 units for the highest standard concentration. The plate should be monitored at 5-minute intervals for approximately 30 minutes.

**Note-3:** If the microplate reader is not capable of reading absorbance greater than the absorbance of the highest standard, perform a second reading at 405 nm. A new standard curve, constructed using the values measured at 405 nm, is used to determine human REDD1 concentration of off-scale samples. The readings at 405 nm should not replace the on-scale readings at 450 nm.

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## Calculations

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Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density. Plot the optical density for the standards versus the concentration of the standards and draw the best curve. The data can be linearized by using log/log paper and regression analysis may be applied to the log transformation. To determine the human REDD1 concentration of each sample, first find the absorbance value on the y-axis and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the x-axis and read the corresponding human REDD1 concentration. If the samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

1. The dose-response curve of this assay fits best to a sigmoidal 4-parameter logistic equation. The results of unknown samples can be calculated with any computer program having a 4-parameter logistic function. It is important to make an appropriate mathematical adjustment to accommodate for the dilution factor.
2. Most microtiter plate readers perform automatic calculations of analyte concentration. The calibration curve is constructed by plotting the absorbance (Y) of calibrators versus log of the known concentration (X) of calibrators, using the 4-parameter function. Alternatively, the logit log function can be used to linearize the calibration curve (i.e. logit of absorbance (Y) is plotted versus log of the known concentration (X) of calibrators).

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## Measurement Range

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The measurement range is 0.2 ng/mL to 12.8 ng/mL. Any sample reading higher than the highest standard should be diluted with Dilution Buffer in higher dilution and re-assayed. Dilution factors need to be taken into consideration in calculating the human REDD1 concentration.

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## Troubleshooting

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1. All samples and standards should be assayed in duplicate, using the protocol described in the **Detailed Protocol**. Incubation times or temperatures significantly different from those specified may give erroneous results.
2. Poor duplicates, accompanied by elevated values for wells containing no sample, indicate insufficient washing. If all instructions in the **Detailed Protocol** were followed accurately, such results indicate a need for washer maintenance.
3. Overall low signal may indicate that desiccation of the plate has occurred between the final wash and addition of Substrate Reagent. Do not allow the plate to dry out. Add Substrate Reagent immediately after wash.

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## Reagent Stability

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All of the reagents included in the MBL Research Product **CircuLex Human REDD1 ELISA Kit** have been tested for stability. Reagents should not be used beyond the stated expiration date. Upon receipt, kit reagents should be stored at 4°C, except the reconstituted Human REDD1 Standard must be stored below -70°C. Coated assay plates should be stored in the original foil bag sealed by the zip lock and containing a desiccant pack.

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## Assay Characteristics

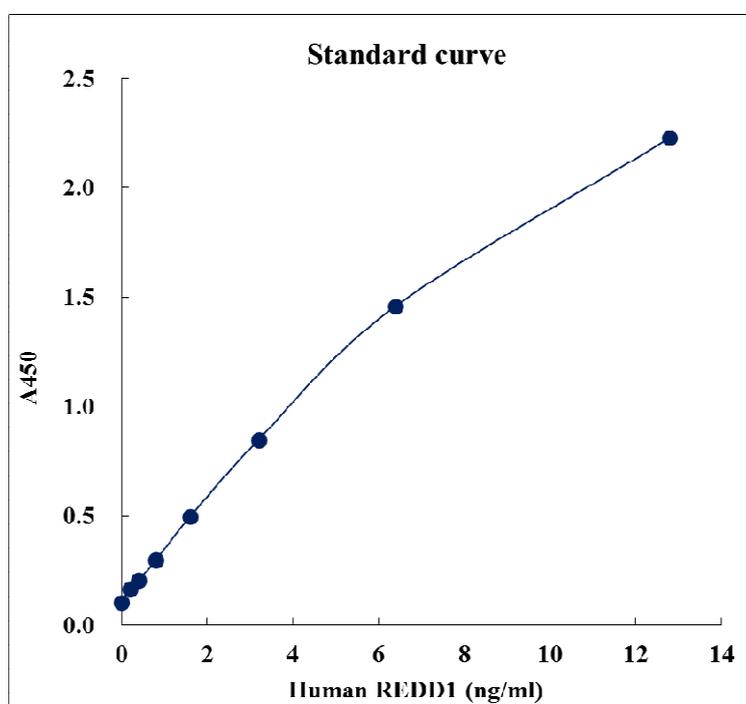
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### 1. Sensitivity

The limit of detection (defined as such a concentration of human REDD1 giving absorbance higher than mean absorbance of blank\* plus three standard deviations of the absorbance of blank: A blank + 3SD blank) is better than 85 pg/mL of sample.

\* Dilution Buffer was pipetted into blank wells.

### Typical Standard Curve



## 2. Precision

### Intra-assay Precision (Precision within an assay)

Three samples\* of known concentration were tested eight times on one plate to assess intra-assay precision.

- Intra-assay (Within-Run, n=8) CV=2.2-4.5%  
Human REDD1 conc. (ng/mL)

\* Sample: Cell lysate

Sample No.	Sample 1	Sample 2	Sample 3
1	2.46	3.56	4.67
2	2.37	3.55	4.40
3	2.33	3.77	4.55
4	2.38	3.81	4.49
5	2.51	3.41	4.59
6	2.36	3.67	4.39
7	2.39	3.79	4.58
8	2.27	3.91	4.60
max.	2.51	3.91	4.67
min.	2.27	3.41	4.39
mean	2.39	3.68	4.53
SD	0.07	0.17	0.10
<b>CV(%)</b>	<b>3.1</b>	<b>4.5</b>	<b>2.2</b>

### Inter-assay Precision (Precision between assays)

Three samples\* of known concentration were tested in four separate assays to assess inter-assay precision.

- Inter-assay (Run-to-Run, n=4) CV=4.3-7.8%  
Human REDD1 conc. (ng/mL)

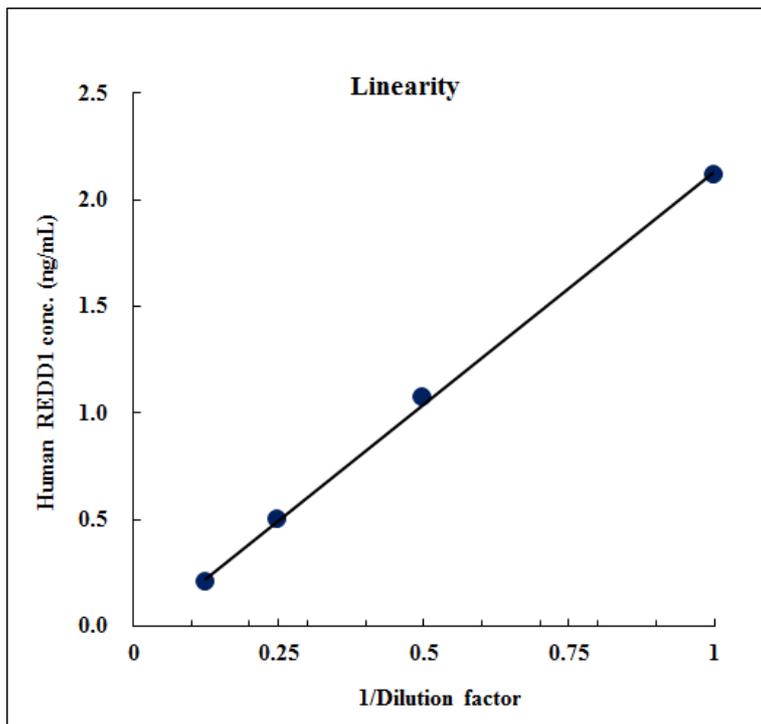
\* Sample: Cell lysate

Sample No.	Sample 1	Sample 2	Sample 3
Assay-1	2.69	3.24	4.23
Assay-2	2.22	3.30	4.25
Assay-3	2.49	3.11	4.64
Assay-4	2.48	3.57	4.36
max.	2.69	3.57	4.64
min.	2.22	3.11	4.23
mean	2.47	3.31	4.37
SD	0.19	0.20	0.19
<b>CV(%)</b>	<b>7.8</b>	<b>5.9</b>	<b>4.3</b>

### 3. Linearity

A sample\* was diluted with Dilution Buffer and assayed after dilution. The neat sample was set to 1.

\* Sample: Cell lysate



### Example of Test Results

Fig.1 Human REDD1 concentrations in cell lysates of HeLa which had been treated with 0-8 mM 2-Deoxy-D-glucose for 6 hours

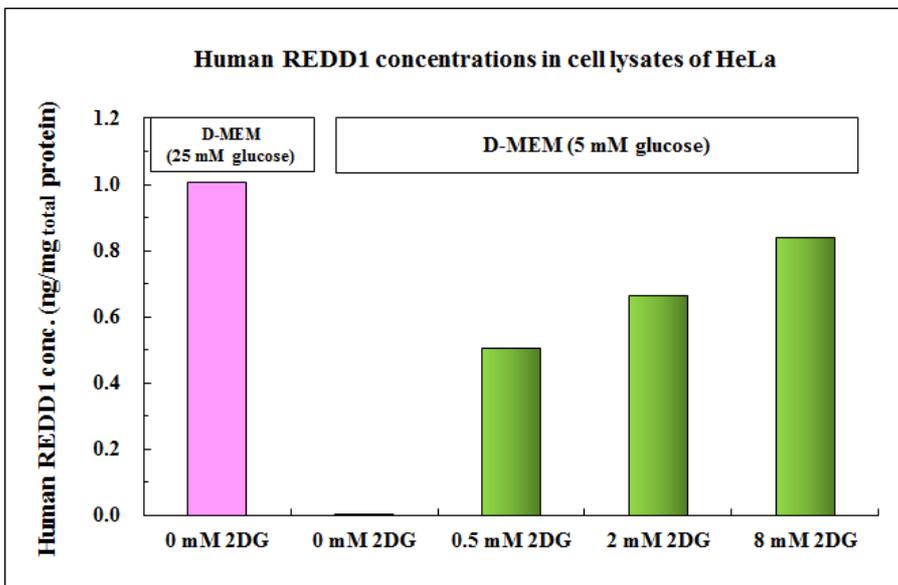
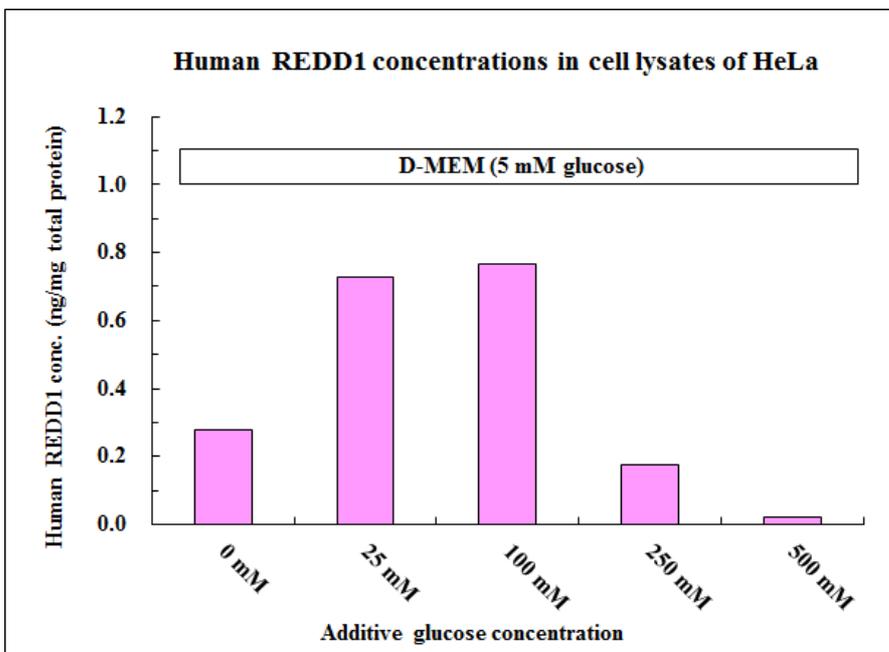


Fig.2 Human REDD1 concentrations in cell lysates of HeLa which had been treated with 0-500 mM glucose for 6 hours



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## References

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2. Shoshani T *et al.* Identification of a novel hypoxia-inducible factor 1-responsive gene, RTP801, involved in apoptosis. *Mol Cell Biol.* 22, 2283-93 (2002)
3. Wang Z *et al.* Dexamethasone-induced gene 2 (dig2) is a novel pro-survival stress gene induced rapidly by diverse apoptotic signals. *J Biol Chem.* 278, 27053-8 (2003)
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6. Brugarolas J *et al.* Regulation of mTOR function in response to hypoxia by REDD1 and the TSC1/TSC2 tumor suppressor complex. *Genes Dev.* 18, 2893-904 (2004)
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## Related Products

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- \* CycLex Cellular UV DNA-Damage Detection Kit: Cat# CY-1141
- \* CycLex Histone H2A.X Phosphorylation Cellular ELISA: Cat# CY-1143
- \* CycLex AMPK Kinase Assay Kit: Cat# CY-1182
- \* CycLex Phospho-p53 Ser46 ELISA Kit: Cat# CY-7050
- \* CycLex Phospho-p53 Ser392 ELISA Kit: Cat# CY-7051
- \* CycLex Phospho-PKR Thr451 ELISA Kit: Cat# CY-7054
- \* CircuLex Human TXNIP ELISA Kit: Cat# CY-8090
- \* CircuLex Human UCHL1 ELISA Kit: Cat# CY-8092
- \* CircuLex Mouse UCHL1 ELISA Kit: Cat# CY-8093
- \* CircuLex Human DJ-1/PARK7 ELISA Kit Ver.2: Cat# CY-9050V2

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