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MONOCLONAL ANTIBODY				
Anti-HVEM				
Code No. K0031-3	Clone 122	Subclass Mouse IgG1 κ	Quantity 100 μg	Concentration 1 mg/mL

**BACKGROUND:** Herpes virus entry mediator (HVEM) is a member of the tumor necrosis factor receptor (TNFR) superfamily that has a role in herpes simplex virus entry, in T cell activation and in tumor immunity. HVEM is constitutively expressed on CD4<sup>+</sup> and CD8<sup>+</sup> T cells, CD19<sup>+</sup> B cells, CD14<sup>+</sup> monocytes, neutrophils and dendritic cells (DCs). The cytoplasmic region of HVEM binds to several members of the TNFR-associated factor (TRAF) family, namely, TRAF1, TRAF2, TRAF3 and TRF5, but not to TRAF4 or TRAF6, and that it activates transcription factors NF-κB and AP-1. The co-expression of glycoprotein D (gD) and HVEM results in the inhibition of the NF-κB activation that is induced by the HVEM overexpression.

- **SOURCE:** This antibody was purified from hybridoma (clone 122) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell Sp2/0 with Balb/c mouse splenocyte immunized with the recombinant HVEM.
- **FORMULATION:** 100 µg IgG in 100 µL volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.
- **STORAGE:** This antibody solution is stable for one year from the date of purchase when stored at -20°C.

**REACTIVITY:** This antibody reacts with human HVEM antigen on Flow cytometry.

# **APPLICATIONS:**

<u>Western blotting;</u> Not tested <u>Immunoprecipitation;</u> Not tested <u>Immunohistochemistry;</u> Not tested <u>Immunocytochemistry;</u> Not tested <u>Flow cytometry;</u> 10-20 µg/mL (final concentration)

Detailed procedure is provided in the following **PROTOCOLS**.

## **SPECIES CROSS REACTIVITY:**

Species	Human	Mouse	Rat
Cell	THP-1	Not Tested	Not Tested
Reactivity on FCM	+		

## **INTENDED USE:**

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### **REFERENCES:**

- 1) Compaan, D. M., et al., J. Biol. Chem. 280, 39553-39561 (2005)
- 2) Gonzalez, L. C., et al., PNAS 102, 1116-1121 (2005)
- 3) Jung, H.W., et al., Exp. Mol. Med. 35, 501-508 (2003)
- 4) La, S., et al., Mol. Cells 14, 398-403 (2002)
- 5) Tamada, K., et al., J.Immunol. 164, 4105-4110 (2000)
- 6) Zhai, Y., et al., J. Clin. Invest. 102, 1142-1151 (1998)
- 7) Kwon, B.S., et al., J. Biol. Chem. 272, 14272-14276 (1997)

Clone 122 is used in the reference 1) - 3).

## **RELATED PRODUCTS:**

K0031-4	FITC labeled Anti-HVEM (122)
K0029-3	CD137 (4B4-1)
K0030-3	CD137L (5F4)
K0030-4	FITC labeled CD137L (5F4)
K0039-3	Anti-TNF-R1 (H398)
K0039-4	FITC labeled Anti-TNF-R1 (H398)
K0040-3	Anti-TNF-R2 (80M2)
K0040-4	FITC labeled Anti-TNF-R2 (80M2)
M031-3	Anti-TRADD (3E11)
D113-3	Anti-Human TNF-α (#1)
D114-3	Anti-Human TNF- $\beta$ (lymphotoxin) (#1)
D125-3	Anti-OX40 (W4-3)
D126-3	Anti-OX40L (TAG-34)
D200-3	Anti-Human BAFF/BlyS (1D6)
D200-4	FITC labeled Anti-Human BAFF/BlyS (1D6)
D201-3	Anti-Human BAFF-R/BR3 (8A7)
D222-3	Anti-GITR (DTA-1)
D222-4	FITC labeled Anti-GITR (DTA-1)
D222-5	PE labeled Anti-GITR (DTA-1)
M028-3	Anti-TRAF1 (3D4)
M112-3	Anti-TRAF2 (6F8)
592	Anti-TRAF2 (poly)
M092-3	Anti-TRAF6 (1F8)
597	Anti-TRAF6 (poly)
4842	IMMUNOCYTO Intracellular TNF-α Detection Kit

# **PROTOCOLS:**

### Flow cytometric analysis for floating cells

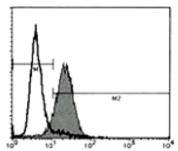
We usually use Fisher tubes or equivalents as reaction tubes for all step described below.

- 1) Wash the cells 3 times with washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.1% NaN<sub>3</sub>].
- 2) Resuspend the cells with washing buffer ( $5x10^6$  cells/mL).

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- 3) Add 50  $\mu$ L of the cell suspension into each tube, and centrifuge at 500 x g for 1 minute at room temperature (20~25°C). Remove supernatant by careful aspiration.
- 4) Add 10  $\mu$ L of normal goat serum containing 1 mg/mL normal human IgG and 0.1% NaN<sub>3</sub> to the cell pellet after tapping. Mix well and incubate for 5 minutes at room temperature.
- 5) Add 40  $\mu$ L of anti-HVEM (122) (25  $\mu$ g/mL) diluted with the washing buffer. Mix well and incubate for 30 minutes at room temperature.
- 6) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 7) Add 30  $\mu$ L of 1:40 FITC conjugated anti-mouse IgG (MBL; code no. IM-0819) diluted with the washing buffer. Mix well and incubate for 15 minutes at room temperature.
- 8) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 9) Resuspend the cells with 500  $\mu$ L of the washing buffer and analyze by a flow cytometer.

(Positive control for Flow cytometry; THP-1)



Flow cytometric analysis of HVEM expression on THP-1 cells. Open histogram indicates the reaction of lsotypic control to the cells. Shaded histogram indicates the reaction of K0031-3 to the cells.

#### Flow cytometric analysis for whole blood cells

We usually use Falcon tubes or equivalents as reaction tubes for all step described below.

- 1) Add 20  $\mu$ L of anti-HVEM monoclonal antibody (122) (50  $\mu$ g/mL) diluted with the washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.1% NaN<sub>3</sub>] into each tube.
- 2) Add 50  $\mu$ L of whole blood into each tube. Mix well, and incubate for 30 minutes at room temperature (20~25°C).
- 3) Add 1 mL of washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 4) Add 30  $\mu$ L of 1:100 FITC conjugated anti-mouse IgG (MBL; code no. IM-0819) diluted with washing buffer. Mix well and incubate for 15 minutes at room temperature.
- 5) Add 1 mL of washing buffer followed by centrifugation at

500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.

- 6) Lyse with OptiLyse C (for analysis on Beckman Coulter instruments) or OptiLyse B (for analysis on BD instruments), using the procedure recommended in the respective package inserts.
- 7) Add 1 mL of  $H_2O$  to each tube and incubate for 10 minutes at room temperature.
- 8) Centrifuge at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 9) Add 1 mL of washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 10) Resuspend the cells with 500  $\mu$ L of the washing buffer and analyze by a flow cytometer.