For Research Use Only. Not for use in diagnostic procedures.



# Anti-CLOCK mAb

CODE No.	D349-3
CLONALITY	Monoclonal
CLONE	CLSP4
ISOTYPE	Mouse IgG1 κ
QUANTITY	100 μL, 1 mg/mL
SOURCE	Purified IgG from hybridoma supernatant
IMMUNOGEN	MBP-Ser/Pro-rich region (Ser <sup>377</sup> -Glu <sup>556</sup> ) of mouse Clock
FORMULATION	PBS containing 50% Glycerol (pH 7.2). No preservative is contained.
STORAGE	This antibody solution is stable for one year from the date of purchase when stored at $-20^{\circ}$ C.

### **APPLICATIONS-CONFIRMED**

Western blotting	$1 \ \mu g/mL$ for chemiluminescence detection system
Immunoprecipitation	2 μg/sample
Immunohistochemistry	Can be used.

#### **APPLICATION-REPORTED**

	<u>C</u>	hromatin	Immuno	preci	pitation	(ChIP)	Reference 2)
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#### SPECIES CROSS REACTIVITY on WB

Species	Human	Mouse	Rat	Hamster
Samples	U2OS	Liver nuclear extract, NIH/3T3	Not tested	Not tested
Reactivity	+	+		

Entrez Gene ID 9570 (Human), 12753 (Mouse)

**REFERENCES** 1) Tsurudome, Y., *et al., Sci. Rep.* **8**, 9072 (2018) [WB]

2) Yoshitane, H., et al., Mol. Cell Biol. 34, 1776-1787 (2014) [ChIP]

3) Yoshitane, H., et al., Mol Cell Biol. 29, 3675-3686 (2009) [IP]

## **RELATED PRODUCTS**

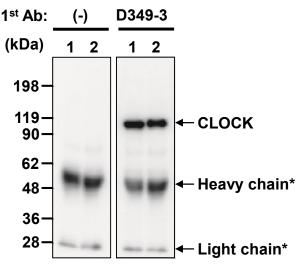
For more information, please visit our web site https://ruo.mbl.co.jp/.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

#### **SDS-PAGE & Western blotting**

- 1) Mix 10  $\mu$ L of Mouse liver nuclear extract with 10  $\mu$ L of Laemmli's sample buffer.
- Boil the sample for 3 min. and centrifuge. Load 20 µL of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel (8% acrylamide) for electrophoresis.
- 3) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 300 mA for 1 hr. in a wet transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacturer's manual for precise transfer procedure.
- 4) To reduce nonspecific binding, soak the membrane in 1% skimmed milk (in TBS, pH 7.2) for 1 hr. at room temperature.
- 5) Incubate the membrane with primary antibody diluted with 1% skimmed milk (in TBS, pH 7.2) as suggested in the **APPLICATION** for 2 hr. at room temperature or overnight at 4°C. (The concentration of antibody will depend on the conditions.)
- 6) Wash the membrane 3 times for 2 min., 5 min. and 10 min. each with 1% skimmed milk (in TBS, pH 7.2).
- 7) Incubate the membrane with 1:5,000 Anti-IgG (Mouse) pAb-HRP (MBL; code no. 330) diluted with 1% skimmed milk (in TBS, pH 7.2) for 2 hr. at room temperature or overnight at 4°C.
- 8) Wash the membrane 3 times for 2 min., 5 min. and 10 min. each with TBS-T [0.05% Tween-20 in TBS].
- 9) Wash the membrane 1 time for 2 min. with TBS.
- 10) Wipe excess buffer on the membrane, and then incubate it with appropriate chemiluminescence reagent for 1 min. Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.
- 11) Expose to an X-ray film in a dark room for 3 min. Develop the film as usual. The condition for exposure and development may vary.

(Positive controls for Western blotting; Mouse liver nuclear extracts)



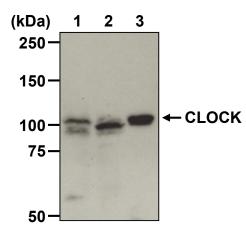
\*The heavy/light chains derived from IgG in the samples. (These bands are detected depending on a sample.)

#### Western blot analysis of mouse CLOCK from liver nuclear extracts

Lane 1: ZT6 (zeitgeber time; 6 h) Lane 2: ZT18 (zeitgeber time; 18 h)

Immunoblotted with Anti-CLOCK (Mouse) mAb (D349-3)

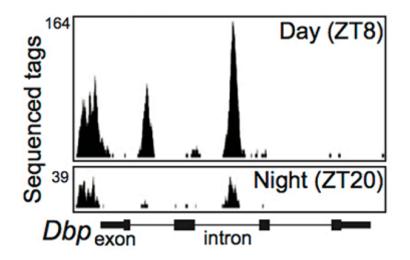
Data were kindly provided by Mr. Kentaro Hirose, Dr. Hikari Yoshitane, and Dr. Yoshitaka Fukada. (Department of Biophysics and Biochemistry, Graduate School of Science, University of Tokyo)



#### Western blot analysis of CLOCK Lane 1: NIH/3T3 Lane 2: U2OS Lane 3: Mouse liver nuclear extract, ZT6 (zeitgeber time; 6 h)

Immunoblotted with Anti-CLOCK (Mouse) mAb (D349-3)

## Chromatin Immunoprecipitation (under evaluation)



## Sequencing analysis of immunoprecipitated DNA using Dbp locus

Sample: Mouse liver nuclear fraction

Antibody: Anti-CLOCK (Mouse) mAb (D349-3; clone CLSP4), 1  $\mu$ g/20  $\mu$ L protein G agarose Sequencing: Genome Analyzer IIx (Illumina)

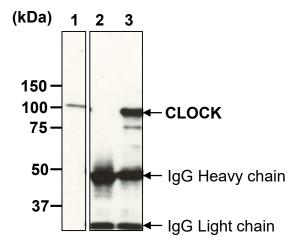
Data were kindly provided by Dr. Hikari Yoshitane, and Dr. Yoshitaka Fukada. (Department of Biophysics and Biochemistry, Graduate School of Science, University of Tokyo)

Reference: Yoshitane, H., et al., Mol. Cell Biol. 34, 1776-1787 (2014) [PMID: 24591654]

## **Immunoprecipitation**

- Add 30 μL of 50% protein G agarose beads slurry resuspended in 100 μL of ice-cold IP buffer [20 mM HEPES-NaOH (pH 7.8), 5.5 mM NaCl, 1 mM EDTA, 6.5% glycerol, 1.5% Triton X-100, 1 mM DTT, 50 mM NaF, 1 mM Na<sub>3</sub>VO<sub>4</sub>] containing appropriate protease inhibitors into the 50 μL of Mouse liver nuclear extract. Incubate it at 4°C with rotating for 30 min.
- 2) Centrifuge the tube at 12,000 x g for 5 min. at 4°C and transfer the supernatant to another tube (precleared sample).
- Add primary antibody as suggested in the APPLICATIONS to the 150 μL of precleared sample (prepared sample from step 2). Incubate with gentle agitation for 1 hr. at 4°C.
- 4) Mix 30 µL of 50% protein G agarose beads slurry into the tube. Incubate with gentle agitation for 1 hr. at 4°C.
- 5) Centrifuge the tube at 2,500 x g for 10 seconds and discard the supernatant.
- 6) Resuspend the agarose with 1 mL of IP buffer.
- 7) Centrifuge the tube at 2,500 x g for 10 seconds and discard the supernatant.
- 8) Repeat steps 5)-7) 4 times. Wash the beads 4 times with 1 mL of IP buffer.
- 9) Resuspend the beads in 20  $\mu L$  of Laemmli's sample buffer, boil for 5 min. and centrifuge.
- 10) Load 20 µL of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel (7.5% acrylamide) for electrophoresis.
- 11) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm<sup>2</sup> for 1 hr. in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacturer's manual for precise transfer procedure.
- 12) To reduce nonspecific binding, soak the membrane in 1% skimmed milk (in PBS, pH 7.2) overnight at 4°C.
- 13) Incubate the membrane with 1 μg/mL of Anti-CLOCK (Mouse) mAb (MBL; code no. D333-3) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hr. at room temperature. (The concentration of antibody will depend on the conditions.)
- 14) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 min. x 3 times).
- 15) Incubate the membrane with 1:10,000 Anti-IgG (Mouse) pAb-HRP (MBL; code no. 330) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hr. at room temperature.
- 16) Wash the membrane with PBS-T (5 min. x 3 times)
- 17) Wipe excess buffer on the membrane, then incubate it with appropriate chemiluminescence reagent for 1 min. Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.
- 18) Expose to an X-ray film in a dark room for 10 min. Develop the film as usual. The condition for exposure and development may vary.

(Positive controls for Immunoprecipitation; Mouse liver nuclear extracts)



### Immunoprecipitation of mouse CLOCK from liver nuclear extracts

- Sample: Mouse liver nuclear extract (ZT6)
- Lane 1: Input (precleared with protein G)
- Lane 2: IP with Mouse IgG1 (M075-3)

Lane 3: IP with Anti-CLOCK (Mouse) mAb (D349-3; clone CLSP4)

Immunoblotted with Anti-CLOCK (Mouse) mAb (D333-3; clone CLSP3)