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# For Research Use Only. Not for use in diagnostic procedures.



## Anti-Pseudouridine mAb

**CODE No.** D347-3

CLONALITYMonoclonalCLONEAPU-6ISOTYPEMouse IgG1 κQUANTITY100 μL, 1 mg/mL

**SOURCE** Purified IgG from hybridoma supernatant

IMMUNOGEN KLH-conjugated pseudouridine

**REACTIVITY** This clone reacts with pseudouridine  $(\Psi)$  containing RNA. Please see the references for more

details.

**FORMULATION** PBS containing 50% glycerol. No preservative is contained.

**STORAGE** This antibody solution is stable for one year from the date of purchase when stored at -20°C.

#### APPLICATIONS-CONFIRMED

Immunohistochemistry 1 μg/mL (paraffin section)

Immunocytochemistry 1 μg/mL

RNA immunoprecipitation Not recommended

#### APPLICATIONS-REPORTED

ELISA Reference 2) and 3) Immuno-Northern blotting Reference 1)

**REFERENCES** 1) Mishima, E., et al., *PLoS One* **10**, e0143756 (2015) [Immuno-Northern blotting]

2) Masuda, M., et al., Cancer 72, 3571-3578 (1993) [IHC-p, ELISA]

3) Itoh, K., et al., Clin. Chim. Acta 181 305-315 (1989) [ELISA]

4) Itoh, K., et al., Tohoku J. Exp. Med. 168, 329-331 (1992)

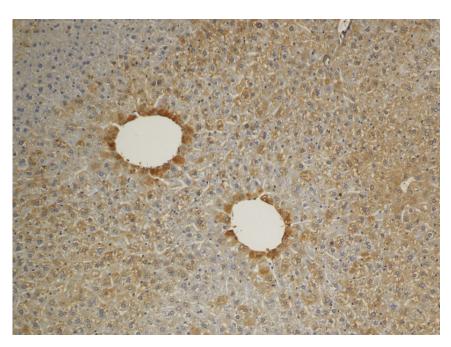
For more information, please visit our web site https://ruo.mbl.co.jp/

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

#### Immunohistochemistry for formalin fixed paraffin-embedded section

- 1) Deparaffinize the section with Xylene (5 min. x 3).
- 2) Wash the slide with Ethanol (5 min. x 3).
- 3) Wash the slide with PBS (5 min. x 3).
- 4) Remove the slide from PBS and inactivate endogenous peroxidase with 3% H<sub>2</sub>O<sub>2</sub> in PBS for 5 min.
- 5) Wash the slide with PBS (5 min. x 3).
- 6) Remove the slide from PBS, wipe gently around the section and incubate with blocking buffer [20 mM HEPES/1% BSA/135 mM NaCl] for 10 min. at room temperature to block non-specific staining. Do not wash.
- 7) Tip off the blocking buffer, wipe gently around each section and incubate with primary antibody diluted with blocking buffer as suggested in the **APPLICATIONS** (The concentration of antibody will depend on the conditions.) for 1 hr. at room temperature.
- 8) Wash the slide with PBS (5 min. x 3).
- 9) Wipe gently around the section and incubate with Histostar<sup>TM</sup> (Ms + Rb) (MBL, code no. 8460) for 30 min. at room temperature.
- 10) Wash the slide with PBS (5 min. x 3).
- 11) Visualize by reacting for 5 min. with Histostar<sup>™</sup> DAB Substrate Solution (MBL, code no. 8469). \*DAB is a suspect carcinogen and must be handled with care. Always wear gloves.
- 12) Wash the slide in water for 5 min.
- 13) Counterstain in hematoxylin for 1 min., wash the slide 3 times in water for 5 min. each, and then immerse the slide in PBS for 5 min.
- 14) Dehydrate by immersing in Ethanol 3 times for 5 min. each, followed by immersing in Xylene 3 times for 5 min. each. Now ready for mounting.

(Positive control for Immunohistochemistry; Mouse liver ischemia model)



#### Immunohistochemistry in mouse liver ischemia model

Brown: Anti-Pseudouridine mAb (MBL, code no. D347-3)

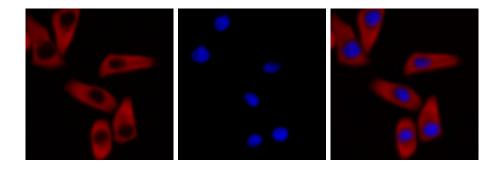
Blue: Hematoxylin

The sample was kindly provided by Dr. Takaaki Abe. (Division of Nephrology, Endocrinology, and Vascular Medicine, Tohoku University Graduate School of Biomedical Engineering)

#### **Immunocytochemistry**

- 1) Spread cells on a glass chamber slide, then incubate in a CO<sub>2</sub> incubator overnight.
- 2) Remove the culture supernatant by careful aspiration.
- 3) Wash the slide twice with PBS.
- 4) Fix the cells with 4% paraformaldehyde/PBS for 20 min. at room temperature (20~25°C).
- 5) Wash the slide 3 times with PBS.
- 6) Permeabilize the cells with 0.5% Triton X-100/PBS for 10 min. at room temperature.
- 7) Wash the slide twice with PBS.
- 8) Block the cells with blocking buffer (1% BSA/PBS) for 1 hr. at room temperature.
- 9) Tip off the blocking buffer and incubate the cells with the primary antibody diluted with blocking buffer as suggested in the **APPLICATIONS** for 1 hr. at room temperature. (Optimization of antibody concentration or incubation condition is recommended if necessary.)
- 10) Wash the slide with 0.05% Tween-20/PBS (5 min. x 3).
- 11) Incubate the cells with 1:1,000 Alexa Fluor® 594 Goat Anti-Mouse IgG (Thermo Fisher Scientific, code no. A-11032) diluted with blocking buffer for 1 hr. at room temperature in dark chamber.
- 12) Wash the slide with 0.05% Tween-20/PBS (5 min. x 3).
- 13) Counterstain with DAPI for 5min. and observe the slide using fluorescent microscopy.

(Positive control for Immunocytochemistry; HeLa)



### Immunocytochemistry in HeLa cells

Red: Anti-Pseudouridine mAb (MBL, code no. D347-3)

Blue: DAPI