For Research Use Only. Not for use in diagnostic procedures.



MONOCLONAL ANTIBODY

Anti-Flavocytochrome b₅₅₈ (Human) mAb

Code No.CloneSubclassQuantityConcentrationD162-37D5Mouse IgG1100 μL1 mg/mL

BACKGROUND: The NADPH oxidase multicomponent enzyme that transfers electrons from NADPH to O₂ to generate superoxide (O₂-), a key part of the phagocytic or neutrophilic respiratory burst response. Flavocytochrome b₅₅₈ is the catalytic component of the phagocyte NADPH oxidase. It is a transmembrane heterodimer composed of a large glycoprotein, gp91^{phox} (PHagocyte OXidase) and a smaller protein, p22^{phox}. Upon cell stimulation, flavocytochrome b558 assembles with p67^{phox}, p47^{phox}, and the GTP-binding protein Rac and becomes activated to generate O2-. Mutations in gp91^{phox}, p22^{phox}, or other components of the NADPH oxidase can result in chronic granulomatous disease, which is associated with significant morbidity and mortality due to a predisposition to recurrent bacterial and fungal infections.

SOURCE: This antibody was purified from hybridoma (clone 7D5) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell Sp2/0 with Balb/c mouse splenocyte immunized with the human cytochrome b rich fraction.

FORMULATION: 100 μg IgG in 100 μL volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at -20°C.

REACTIVITY: This antibody reacts with Flavocytochrome b₅₅₈ flow cytometry.

APPLICATIONS:

<u>Western blotting</u>; Not tested <u>Immunoprecipitation</u>; Not tested <u>Immunocytochemistry</u>; Not tested <u>Immunohistochemistry</u>; Not tested

Flow cytometry; 5 µg/mL (final concentration)

Detailed procedure is provided in the following **PROTOCOL**.

INTENDED USE:

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RELATED PRODUCTS:

Please visit our website at https://ruo.mbl.co.jp/.

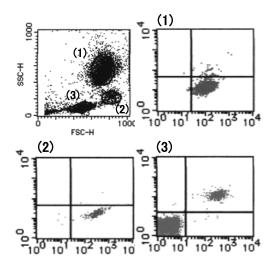
SPECIES CROSS REACTIVITY:

Species	Human	Mouse	Rat
Cells	Lymphocyte Monocyte Granulocyte	Not tested	Not tested
Reactivity on FCM	+		

REFERENCES:

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- 3) Li, X. J., et al., J. Leukoc. Biol. 81, 238-249 (2007) [FCM]
- 4) Yamauchi A, et al., Microbiol. Immunol. 45: 249-57 (2001)
- 5) Burritt, J. B., et al,. J. Biol. Chem. 276: 2053-2061 (2001)
- 6) Yu, L., et al., Blood 94: 2497-2504 (1999)
- 7) Yu, L., et al., PNAS 95: 7993-7998 (1998)
- 8) Radeke, H. H., et al., J. Biol. Chem. 266: 21025-21029 (1991)
- 9) Verhoeven, A., et al., Blood **73**: 1686-1694 (1989)
- 10) Nakamura, M., et al., Blood 69:1404-1408 (1987)

Clone 7D5 is used in these references.



Flow cytometric analysis of Flavocytochrome b₅₅₈ expression on Granulocytes (1), Monocytes (2) and Lymphocytes (3). The staining intensity of D162-3 is shown in the horizontal axis withCD19-FITC staining on the vertical axis.

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The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

PROTOCOL:

Flow cytometric analysis for whole blood cells

We usually use Falcon tubes or equivalents as reaction tubes for all step described below.

- 1) Add 20 μ L of the primary antibody as suggested in the **APPLICATIONS** diluted with the washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.09% NaN₃] into each tube.
- 2) Add 50 μ L of whole blood into each tube. Mix well, and incubate for 30 minutes at room temperature (20~25°C).
- 3) Add 1 mL of washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 4) Add 20 μL of PE conjugated anti-mouse IgG (H+L) antibody diluted with washing buffer. Mix well and incubate for 15 minutes at room temperature.
- 5) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 6) Add 20 μL of FITC conjugated anti-human CD 19 antibody. Mix well and incubate for 15 minutes at room temperature.
- 7) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 8) Lyse with OptiLyse C (for analysis on Beckman Coulter instruments) or OptiLyse B using the procedure recommended in the respective package inserts.
- 9) Add 1 mL of H₂O to each tube and incubate for 10 minutes at room temperature.
- 10) Centrifuge at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 11) Add 1 mL of washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 12) Resuspend the cells with 500 μL of the washing buffer and analyze by a flow cytometer

(Positive controls for flow cytometry: Human granulocyte, monocyte and lymphocyte)